
COMPATIBILITY STUDIES OF OLANZAPINE (API) WITH PLGA 750S POLYMER

Sudhakar Jha,

Research Scholar, Dept. of Pharmacy, Sunrise University, Alwar, Rajasthan

Dr. Vachaspati Dubey,

Research Supervisor, Dept. of Pharmacy, Sunrise University, Alwar, Rajasthan

ABSTRACT

The microspheres were characterized by in-vitro dissolution and other Physiochemical methods. A simulation of multiple dosing at weekly or 15 -day regimen revealed pulsatile behavior for all formulations with steady state may be achieved by the second dose. The current studies was design and evaluate biodegradable PLGA microspheres for sustained or extended release, with primary goal of avoiding combination of oral therapy for the treatment of schizophrenia. PLGA copolymers 75:25 was used to prepare four microsphere formulations of anti-psychotic drug Olanzapine. Overall, the in-vitro study of Formulations 001, 002, 003, or 004 will eliminate the need for combination oral therapy and reduce time to achieve steady state, with a smaller washout period upon duration of therapy. Results of this study prove the suitability of using PLGA 75:25 copolymers of different composition and molecular weight to produce sustained or extended release formulations that can enhance pharmacological effectiveness for anti-psychotic intra-muscular administration of Olanzapine.

Keywords: *sustained release, PLGA microspheres, Olanzapine*

INTRODUCTION

With high patient compliance, microspheres are now regarded as a dependable and efficient release mechanism that lowers doses, frequency of dosing, and the risk of dosage dumping (2). For example, polyesters employed in microspheres have the advantage of employing water as a vehicle to generate suspensions which allows consistent release of desired medicine. Microspheres prepared by natural or synthetic polymers provides sustained release. It has been established that a medication's physicochemical characteristics, main burst, and excipients all have a direct impact on how quickly the drug leaves the microsphere³. Injectable microsphere systems were developed in late 70's. Poly (lactide) and/or poly (lactide-co-glycolide) was used to encapsulate an ester of norethisteron called NET via oil-in-water emulsion/solvent evaporation process. These microcrystals of NET were encapsulated into microspheres of PLA and PLGA. Since then PLGA has been the first choice of polymer for preparation of microspheres to encapsulate APIs using w/o or o/w emulsion/solvent evaporation technique. Poly (ϵ -caprolactone)

is the secondly most accepted polymer for preparation of microspheres to encapsulate APIs. Moreover PLGA, PCL and PLA, diblock copolymers of lactide, ethylene glycol, L-lactide and ϵ -caprolactone, triblock co polymers of caprolactons, lactides and glycolides, have also been used to encapsulate APIs via o/w emulsion / solvent evaporation. ⁽³⁾The composition of the microspheres and the rate at which they degrade have an impact on the release profiles of the API. For instance, the hydrolytic breakdown rate of PGA, PLA, and PCL is in the following order: PGA > PLA >> PCL. In contrast to PLA and PGA, which had inferior permeability in the case of steroidal pharmaceuticals but showed uniform biodegradation, PCL has great permeability in the case of steroidal medications but exhibits sluggish biodegradation. When the system's hydrophilic glycolide component is increased, it has been found that PLGA microspheres breakdown more quickly and release API more quickly. The release patterns of API are further influenced by the polymer's chirality, density of cross-linking, and loading of medicines in microspheres. When the amount of cross-linking is greatly increased, it causes an increase in the barrier density for drug diffusion and a slower release of the API in casein and chitosan microspheres. When unencapsulated medication is more readily available on the surface of microspheres in greater quantities, there is a higher initial burst in microspheres. ⁽³⁾

Materials and Methods

Compatibility of API with polymer:

Compatibility of Olanzapine and polymer PLGA was tested on following parameters.

FTIR Analysis:

FTIR analysis of physical mixture of olanzapine and PLGA was performed on Perkin Elmer FTIR spectrometer.

DSC analysis of physical mixture of Olanzapine and PLGA

DSC analysis of physical mixture of olanzapine and PLGA was performed on Mettler Toredon which is shown below figure 5.6 Approximately 2 mg of API and polymer mixture was weighed in standard aluminum sample holder. In first cycle the temperature sample holder was brought to -10°C . Then the temperature of sample holder was raised at the rate of 10°C per minute up to 60°C . In the Second cycle the temperature sample holder was decreased to -10°C at rate of 10°C per minute. In third cycle temperature of sample holder again raised from -10°C up to 100°C at rate of 10°C per minute. Air flow of 50mL per Minute was maintained throughout the experiment.

RESULT & DISCUSSION

Compatibility of olanzapine (API) with PLGA 750S polymer

FTIR analysis of physical mixture

The physical mixing of olanzapine and PLGA revealed 17 peaks in the FTIR analysis that are characteristic of PLGA and olanzapine. Peak at 2843.95 cm⁻¹ is dedicated to CH₂, CH₃, and asymmetric stretching; peak at 1587.57 cm⁻¹, 1220.17 cm⁻¹ for C=N stretching, 1005.75 cm⁻¹ for aromatic ring deformation, and peak at 971.65 cm⁻¹ for C-S stretching are characteristic of Olanzapine; and peak at 2843.95 cm⁻¹ is dedicated to C=O stretching; these are the characteristics of PLGA and PLGA, respectively. The FTIR demonstrates that the physical mixture of API and polymer does not exhibit any chemical reactions, proving their compatibility.

Table 6.5 : FTIR Interpretation for physical mixture of olanzapine and PLGA polymer

Sr.	Wave Number	Interpretation
1	2843.95	CH ₂ , CH ₃ , CH ₂ asymmetric stretching
2	1751.67	C=O stretching
3	1587.57	C-N stretching
4	1560.61	C-N stretching
5	1455.61	CH ₂ deformation
6	1420.14	CH ₃ deformation
7	1364.98	CH ₂ wagging
8	1284.69	CH ₂ twisting
9	1269.79	CH ₂ twisting
10	1220.17	C=N stretching
11	1180.29	C-H deformation
12	1138.02	Aromatic ring stretching CH deformation
13	1086.22	C-O stretching
14	1044.44	Aromatic ring stretching C-H deformation
15	1005.75	Aromatic ring deformation

16	971.65	C-S stretching
17	851.79	C-H out of plane banding
18	757.62	C-H out of plane deformation
19	711.31	Aromatic ring deformation

DSC analysis of physical mixture of olanzapine and PLGA Polymer 750S

Using the same set of conditions used for the DSC study of Polymer PLGA 750S, the glass transition temperature for the PLGA and Olanzapine mixture was established on Mettler Toledo equipment. Researchers discovered that the glass transition started at 49.44 C and reached its end point at 51.48 C. Olanzapine produces a physical combination without undergoing any chemical reactions, as shown by a comparison with the DSC thermogram of pure polymer PLGA 750S. Also, it is evident from the mixture's DSC that there was no thermal decomposition or chemical reaction in the physical mixture (glass transition temperature of pure PLGA was 48.69 C).

Figure 6.3: DSC Thermogram of physical mixture of olanzapine and PLGA Polymer 750S

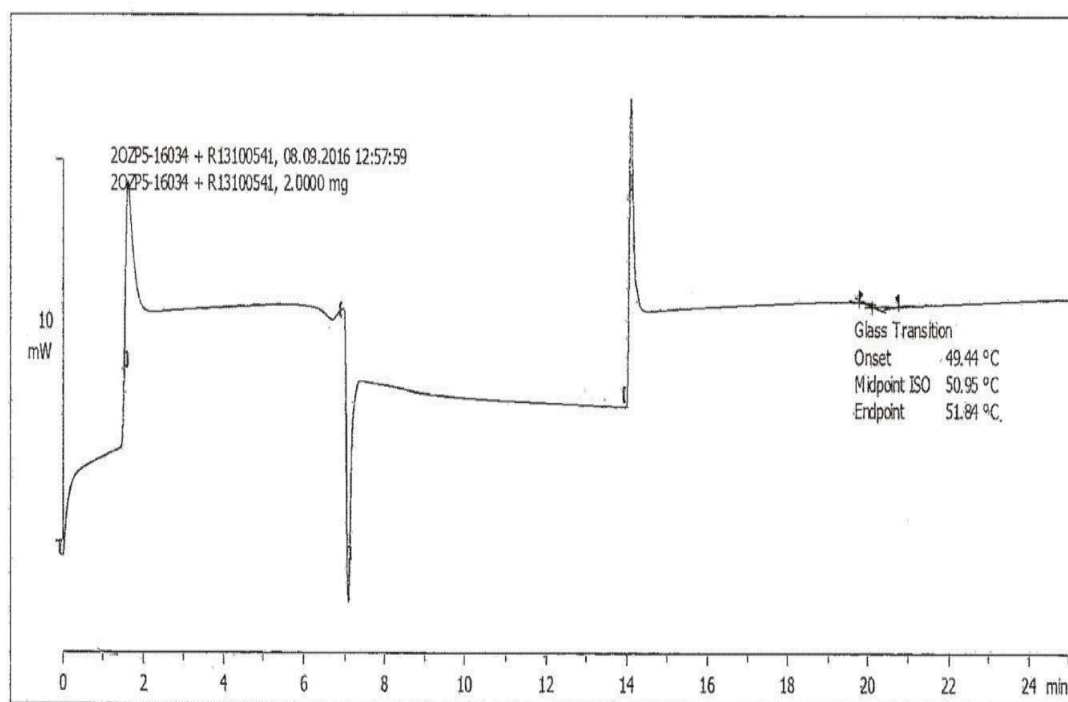


Figure 6.3 : DSC Thermogram of physical mixture of olanzapine and PLGA Polymer 750S

CONCLUSIONS

Olanzapine is a second-generation analytical antipsychotic medication. It is used to treat mild to severe mania associated with bipolar illnesses as well as schizophrenia. The principal oral dose, which can be given as a single therapeutic agent or as part of a combination therapy, is 10 mg per day. Degradation of PLGA depends on percentage of glycolic monomer in polymer. Higher the quality of glycolic acid lower is the time for degradation. Hence PLGA with 75:25 lactic acid : Glycolic acid ratio will take higher time for degradation in comparison to 50:50 lactic acid to glycolic acid ratio. Keeping in mind required properties of polymer PLGA 755 (lactic acid: Glycolic 75:25) was selected in present study to prepare microsphere of olanzapine.

REFREANCES

1. Banker, G. S. and Rhodes, C.T., “Modern Pharmaceutics: Fourth Edition, Revised and Extended”, Marcel Dekker Inc., New York, Basel, 2002, Chapter 15, Page.
2. D’Souza, S.S. and Deluca, P.P., “Development of a Dialysis *In vitro* Release Method for Biodegradable Microspheres”, AAPS Pharmaceutical Science and Technology, 2005, Issue 6(2), Article 42.
3. Wu, Linfeng, “Long Acting Injectable Hormonal Dosage Forms for Contraception”, Pharmaceutical Research, 2015, Issue 32(2), Page 180- 219.
4. Wise, Donald L., “Handbook of Pharmaceutical Controlled Release Technology”, Marcel Dekker Inc., New York, Basel, 2000, Chapter 13, Page271-275.
5. Wise, Donald L., “Handbook of Pharmaceutical Controlled Release Technology”, Marcel Dekker Inc., New York, Basel, 2000, Chapter 15, Page301-309.
6. Wise, Donald L., “Handbook of Pharmaceutical Controlled Release Technology”, Marcel Dekker Inc., New York, Basel, 2000, Chapter 16, Page329-338.
7. Ye M, Kim S, Park K. Issues in long-term protein delivery using biodegradable microparticles. *Journal of Controlled Release*, 2010, 146(2): 241–260.
8. Morlock M, Koll H, Winter G, Kissel T. Microencapsulation of Rherythropoietin, using biodegradable poly(D,L-lactide-co-glycolide): Protein stability and the effects of stabilizing excipients. *European Journal of Pharmaceutics and Biopharmaceutics*, 1997, 43(1): 29–36.
9. Meinel L, Illi O E, Zapf J, Malfanti M, Merkle H P, Gander B. Stabilizing insulin-like growth factor-I in poly(D,L-lactide-coglycolide) microspheres. *Journal of Controlled Release*, 2001, 70 (1-2): 193–202.
10. Kang J, Wu F, Cai Y P, Xu MX, He M, Yuan WE. Development of recombinant human growth hormone (RhGH) sustained-release microspheres by a low temperature aqueous phase/aqueous phase emulsion method. *European Journal of Pharmaceutical Sciences*, 2014, 62: 141–147.

11. Hong X Y, Wei L M, Ma L Q, Chen Y H, Liu Z G, Yuan W. Novel preparation method for sustained-release PLGA microspheres using water-in- oil-in-hydrophilic-oil-in-water emulsion. *International Journal of Nanomedicine*, 2013, 8: 2433–2441.
12. Kumar R, Palmieri MJ Jr. Points to consider when establishing drug product specifications for parenteral microspheres. *AAPS Journal*, 2010, 12(1): 27–32
13. Toguchi H. Sterility assurance of microspheres. *Journal of Controlled Release*, 1999, 62(1-2): 51–55.
14. Wong J, Brugger A, Khare A, Chaubal M, Papadopoulos P, Rabinow B, Kipp J, Ning J. Suspensions for intravenous (IV) injection: A review of development, preclinical and clinical aspects. *Advanced Drug Delivery Reviews*, 2008, 60(8): 939–954
15. Nath S D, Son S, Sadiasa A, Min Y K, Lee B T. Preparation and characterization of PLGA microspheres by the electrospraying method for delivering simvastatin for bone regeneration. *International Journal of Pharmaceutics*, 2013, 443(1-2): 87–94.
16. Zhang M, Ma Y, Li R, Zeng J, Li Z, Tang Y, Sun D. RhBMP-2- loaded Poly(lactic-co-glycolic acid) microspheres fabricated by coaxial electrospraying for protein delivery. *Journal of Biomaterials Science. Polymer Edition*, 2017, 28(18): 2205–2219.
17. Della Porta G, Campardelli R, Cricchio V, Oliva F, Maffulli N, Reverchon E. Injectable PLGA/hydroxyapatite/chitosan microcapsules produced by supercritical emulsion extraction technology: An in vitro study on teriparatide/gentamicin controlled release. *Journal of Pharmaceutical Sciences*, 2016, 105(7): 2164–2172.
18. Falco N, Reverchon E, Della Porta G. Injectable PLGA/hydrocortisone formulation produced by continuous supercritical emulsion extraction. *International Journal of Pharmaceutics*, 2013, 441 (1-2): 589–597.
19. Della Porta G, Campardelli R, Reverchon E. Monodisperse biopolymer nanoparticles by continuous supercritical emulsion extraction. *Journal of Supercritical Fluids*, 2013, 76: 67–73.
20. Campardelli R, Della Porta G, Gomez V, Irusta S, Reverchon E, Santamaria J. Encapsulation of titanium dioxide nanoparticles in PLA microspheres using supercritical emulsion extraction to produce bactericidal nanocomposites. *Journal of Nanoparticle Research*, 2013, 15(10): 1987– 1997.
21. Della Porta G, Falco N, Giordano E, Reverchon E. PLGA microspheres by supercritical emulsion extraction: A study on insulin release in myoblast culture. *Journal of Biomaterials Science. Polymer Edition*, 2013, 24(16):1831–1847.
22. Della Porta G, Nguyen B N, Campardelli R, Reverchon E, Fisher J P. Synergistic effect of sustained release of growth factors and dynamic culture on osteoblastic differentiation of mesenchymal stem cells. *Journal of Biomedical Materials Research. Part A*, 2015, 103(6): 2161–2171.

23. Campardelli R, Della Porta G, Gomez L, Irusta S, Reverchon E, Santamaria J. Au-PLA nanocomposites for photothermally controlled drug delivery. *Journal of Materials Chemistry. B, Materials for Biology and Medicine*, 2014, 2(4): 409–417.
24. Jiang W L, Schwendeman S P. Stabilization of tetanus toxoid encapsulated in PLGA microspheres. *Molecular Pharmaceutics*, 2008, 5(5): 808–817.
25. Liu Z Q, Li X, Xiu B S, Duan C M, Li J X, Zhang X H, Yang X Q, Dai W H, Johnson H, Zhang H Q, et al. A novel and simple preparative method for uniform-sized PLGA microspheres: Preliminary application in antitubercular drug delivery. *Colloids and Surfaces. B, Biointerfaces*, 2016, 145: 679–687.
26. Hung L H, Teh S Y, Jester J, Lee A P. PLGA micro/nanosphere synthesis by droplet microfluidic solvent evaporation and extraction approaches. *Lab on a Chip*, 2010, 10(14): 1820–1825.
27. Wei Y, Wang Y X, Wang L Y, Hao D X, Ma G H. Fabrication strategy for amphiphilic microcapsules with narrow size distribution by premix membrane emulsification. *Colloids and Surfaces. B, Biointerfaces*, 2011, 87(2): 399–408.
28. Qi F, Wu J, Fan Q Z, He F, Tian G F, Yang T Y, Ma G H, Su Z G. Preparation of uniform-sized exenatide-loaded PLGA microspheres as long- effective release system with high encapsulation efficiency and biostability. *Colloids and Surfaces. B, Biointerfaces*, 2013, 112: 492–498
29. Crowley M M, Zhang F, Repka M A, Thumma S, Upadhye S B, Battu S K, McGinity J W, Martin C. Pharmaceutical applications of hot-melt extrusion: Part I. *Drug Development and Industrial Pharmacy*, 2007, 33(9): 909–926.
30. Guo Y, Yang Y, He L, Sun R, Pu C, Xie B, He H, Zhang Y, Yin T, Wang Y, Tang X. Injectable sustained-release depots of PLGA microspheres for insoluble drugs prepared by hot-melt extrusion. *Pharmaceutical Research*, 2017, 34(10): 2211–2222.
31. Bakri S J, Omar A F. Evolution of vitreomacular traction following the use of the dexamethasone intravitreal implant (Ozurdex) in the treatment of macular edema secondary to central retinal vein occlusion. *Journal of Ocular Pharmacology and Therapeutics*, 2012, 28(5): 547–549.
32. Tice T. US Patent 2012/0156304 A1, 2012–06-21.
33. Liu R, Ma G H, Meng F T, Su Z G. Preparation of uniform-sized PLA microcapsules by combining Shirasu Porous Glass membrane emulsification technique and multiple emulsion-solvent evaporation method. *Journal of Controlled Release*, 2005, 103(1): 31–43.
34. Liu R, Ma G H, Wan Y H, Su Z G. Influence of process parameters on the size distribution of PLA microcapsules prepared by combining membrane emulsification technique and double emulsion-solvent evaporation method. *Colloids and Surfaces. B, Biointerfaces*, 2005, 45(3-4): 144–153.

35. Liu R, Huang S S, Wan Y H, Ma G H, Su Z G. Preparation of insulin-loaded PLA/PLGA microcapsules by a novel membrane emulsification method and its release in vitro. *Colloids and Surfaces. B, Biointerfaces*, 2006, 51(1): 30– 38.